



# Ecological Spectra Relation to Digenean Trematode Parasites in Fresh Water Fish *Channa Striatus* (BLOCH)

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## ABSTRACT

Fishes are found adequately in the different water sources of Jaunpur India. Five rivers (Gomti, Sai, Varuna, Pili and Basuhi), Gujarat and different ponds are available here as an aquatic habitat. In research work, the author considers only two sites of river Gomti and two different ponds in district Jaunpur, U.P. They are external as well as internal. In this study, the author focused on digenetic trematode parasites. Digenean trematodes complete their life cycle in two hosts so both are susceptible to the infection. There are considerable changes that occur in the environment with the passage of time. We also know that environmental factors affect the life forms of any place. So it is important to study the different ecological terms in relation to the host and their infectious agents. From ancient times, humans have used a large amount of fish food for survival because they are easily available from different water sources. When human beings eat inadequate cooked fish, they suffer from food-borne trematodiasis. So it is important to know the infection level in the fishes.

**KEYWORDS** :- Parasites, *Channa striatus* & Ecological parameters.

## INTRODUCTION

The property of easy availability of fishes is attracting human beings from ancient times to fulfill the need of their food. Now a days fish culture has become an industry for a country which has adequate water sources. India is the second largest producer of fish with an annual production of 312.8 million tonnes (FAO, 2008). Fish interact with the various levels of food chain and influence the structures of lakes, streams and estuaries since they are usually restricted to particular modes of life related to their food sources and reproductive requirements (Ashade et al., 2013). There are a number of helminth parasites which are

transmitted to human beings only through fish (Gupta, 1959). These parasites use the fish for their shelter and food and destruct every organ resulting in pathogenic effects (Dogiel, 1958). Parasites interfere with the nutrition, metabolism and secretory function of alimentary canal, damage nervous system (Markov, 1961). Parasites do not represent a monophyletic group, they are an assembly of organisms belonging to separate lineages in which a parasitic mode of life has evolved independently (Zrzavy 2001; De Meeus and Renaud 2002) and organ distribution of helminth parasites in *Macrognathus aculeatus* (Khanum et al., 2011). Distribution of helminth parasites in

different size group and organs of fish (D'Silva et al.,2012).However the diversity of parasites within a single host individual is dependent on many factors ,from the season of capture to purely stochastic factors and is not often a reflection of the diversity of the pool of locally available parasite species, we will not focus on this level(Robert Poulin,2004).For instance, infracommunities of intestinal helminths in Eels, *Anguilla anguilla* become saturated with parasite species at values well below the number of parasite species locally available (Kennedy and Guegan 1996).Infracommunities of larval trematodes in gastropod hosts provide an even better illustration :individual snail almost always harbor a single trematode species ,whereas several trematode species can coexist in the same snail population(Kuris and Lafferty 1994;Esch et al.,2001). The spotted murrel,*Channa punctata*( bloch)is regarded as a delicious food fish in India (Shareef et al.,2012).

A survey of metacercarial infections in commonly edible fish and crabs host(Athokpam et al.,2015).Prevalence of helminth parasites infecting *Channa punctatus*(Kundu et al.,2015).An ecological aspects on digenetic trematode parasites of fresh water fish (Chandra et al.,2016) and Presence of digenetic trematode parasites in fresh water carnivorous fishes in Jaunpur (Singh et al.,2018).Prevalence of parasitic infections in the fresh water fishes *Channa punctatus* (Sarmin et al.,2018).It was reported that the unfavourable environmental and ecological conditions caused a variety of fish parasites (Chandra et al., 1987).Also described the relationship between digenetic trematode parasites and cyprinid fish *Catla catla*(Mishra et al.,2019)and occurrence of digenetic trematode parasites in fresh water catfish *Wallago attu*(Singh et al.,2019). Presence of digenetic trematode parasites of cypriniformes fish *Labeo rohita*(Mishra et al.,2019).During study of different research papers and Thesis author found that a little work was done in ecological relation of *Channa striatus* and digenetic trematode parasites in District Jaunpur .In past few years no work has been done on trematode parasites in Jaunpur.Their number is also declined in recent few years .So it is important to focus on the relation of *Channa striatus* and digenetic trematode parasites in ecological terms. In this study author tried to know the

relation of digenetic trematode parasites and *Channa striatus*.

### Materials and Methods:-

(a) Sample Source –Sample were collected from the different site of river Gomti (Kalichabad ghat & Hanuman ghat) and ponds (Banaba pond and Kandharpur pond) in district Jaunpur.

(b)Working time -1July 2020 to 30 June 2021.

(c)Sample collection–Fishes were collected from the given sources with the help of fishing net. Fishermen were also helped with their boats. Fishes of all size, weight and sex were collected alive from the different sites. They were collected in a plastic container and carry to the laboratory.In the laboratory samples poured in the soil jar.

(d)Sample and Digenetic Trematode parasites-*Channa striatus* , the striped snakehead ,is a species of snakehead fish .*Channa striatus* is native to fresh water habitats in South and Southeast Asia .The species can reach up to a meter in length, through because of fishing, this size is rarely found in the world .It has a widespread range covering Southern China, Pakistan ,most of India, Southern Nepal , Bangladesh ,Sri Lanka and most of Southeast Asia. It is an important food in its entire native range and is of considerable economic importance. Adults are dark brown in colour with faint black bands visible across its entire body. Eggs are guarded by both parents.It is common in fresh water plains, where it migrates from rivers and lakes into flooded fields,returning to the permanent water bodies in the dry season, where it survives by burrowing in the mud.It prey frogs, water bugs and smaller fishes(Wik.).



***Channa striatus* (Saur)**

The digenetic trematode parasites have syncytial tegument with two suckers ,one ventral and one oral.They are mostly occur in the digestive system but present throughout the organ system.

(e)Standard length and weight measurement–Samples were measured with the help of measuring board and



weighing machine was used for weight measurement. Standard protocol was followed (Paperna I.1996).

(f) *Examination, Processing of digenetic parasites for identification and statistical formula-* Author put the sample in a dissecting tray .Scissors, forceps and needle were used for opening the internal parts .They dissected darsoventrally and examined by using standard protocol.We removed internal organs like liver ,kidney air-bladder, heart and alimentary canal from the fish body .Then every organs and tissues put separately in a petri dish and washed many times with the tap water and finally filled with 0.70 % saline solution.Ascending grades of alcohol were used for the dehydration .Parasites were collected in test tube filled with 70% ethanol for 24-48 hours.After that counted one by one.Trematodes were stained in aceto-alum carmine.They were mounted in canada balsam and followed by Camera lucida diagrams,then examined under a binocular microscope. They were identified with help of Book Systema Helminthol (Yamaguti S.1954). Different formulas are used to determine the ecological terms proposed (Margolis I. et al., 1982)and given in his concluded note that a variety of terms in addition to intensity and prevalence have been used to refer either specifically or generally to the numbers of parasites occurring in a host or to the number of infected hosts in a sample.

$$\text{Prevalence \%} = \frac{\text{Total no of fish infected}}{\text{Total no. of fish host examined}} \times 100$$

$$\text{Abundance} = \frac{\text{Total no. of parasites recovered}}{\text{Total no. of fish host}}$$

$$\text{Mean Intensity} = \frac{\text{Total no of parasites recovered}}{\text{Total no. of host infected}}$$

$$\text{Index of infection} = \frac{\text{No. of host infected} \times \text{No. of parasites recovered}}{\text{Total host examined}}$$

$$\text{Dominant\%} = \frac{\text{Worm burden monthly} \times 100}{\text{Worm burden annually}}$$

## Results :-

In the monthly observation we found that Prevalence was maximum in the month of July and minimum in the month of December. Abundance was maximum in the month of October and minimum in the month of December. Mean intensity was maximum in the month of October and minimum in the month of December. Dominance was maximum in the month of July and minimum in the month of December. Infestation index was maximum in the month of July and minimum in the month of December.

On the basis of seasonal aspects: Prevalence was maximum in the Rainy season and minimum in the Winter season. Abundance was maximum in the Rainy season and minimum in the Winter season. Mean intensity was maximum in the Rainy season and minimum in the Winter season. Dominance was the maximum in the Rainy season and minimum in the Winter season. Infestation index was maximum in the Rainy season and minimum in the Winter season.

Fig. 1

FAMILY-MASENIIDAE

Masenia jaunpurensis Sp. nov.

Location: Intestine

The body of trematode is extended, spinose, round and hollow fit as a fiddle with restricted front and wide back closures and measures 1.90 long and 0.59 in width. The oral suker is circular, subterminal and measures 0.13x0.14. The ventral sucker is oval, preequatorial, and somewhat littler than oral sucker, middle and measures 0.16 x 0.14. The mouth leads into a short, cylindrical prepharynx which estimates 0.03 x 0.02 long. The pharynx is solid, adjusted which estimates 0.032 x 0.034. Throat little and bifurcates at the separation of 0.33 from the front finish of body into two intestinal caeca which broadens upto the foremost degree of front testis.Excretory bladder cylindrical and excretory pore terminal Testicles postequatorial, oval or adjusted, marginally over lapping, submedian and measures 0.13 x 0.15 foremost testis and 0.14 x 0.15 back testis. Cirrus sac long, cylindrical reaching out from genital pore upto front area of hip bone socket and measures 0.53 long Basal bit is wide while distal end is restricted. Cirrus sac encases whole vesiculum seminalis. Rounded standards prostatica encompassed by prostatic organ cells. Ejaculatory conduit is long, cylindrical with little cirrus. Ovary is three-sided, pretesticular, - submedian,

preequatorial, somewhat covering the privilege ceacum and measures  $0.19 \times 0.15$ . Receptaculum seminis long, club formed lying sidelong to the overy and measures  $0.16 \times 0.06$ . Mehli's organ cells encompass the ootype. Uterus wound, broadening upto back finish of body and rising corresponding to the cirrus sac to open at genital pore. Eggs oval operculated and measures  $0.024 \times 0.011 - 0.022 \times 0.010$ . Genital pore on the dorsal side of oral sucker. Vitelline follicles enormous, stretching out from the center degree of hip bone socket upto, center degree of front testis. The new structure has a place with the sort Masenia Chatterji, of which the accompanying nine species have been depicted from Indian fresh water fishes viz. *M. colcatta* Chatterji, from *Clarias batrachus* \ *M. dayali* Gupta, from *Clarias batrachus*; *M. fossilisi* Gupta, from *Heteropneustes fossilis*; *M. gomtia* Agrawal, from *Mystus vittatus*; *M. ritai* Sircar and Sinha; *M. yamaguti* Agrawal; *M. upeneusi* Gupta and Puri, and *M. chauhani* Maurya and Agrawal, from *rita*.

Fig. 2

#### FAMILY-MASENIIDAE

*Masenia dayali* Gupta, 1955

Location: Intestine

The body of Trematode is lengthened, cylindrical, and spinose with somewhat tight foremost and expansive back closures and measures. 1.60 long and 0.49 in width Oral sucker' channel formed subterminal with twofold circum oral crown of spines and measures  $0.09 \times 0.10$ . The ventral sucker is oval or roundabout, pre-central marginally submedian and measures  $0.11 \times 0.09$ . Prepharynx long, rounded, thin and quantifies 0.13 long. Pharynx very much created, solid, subglobular and measures  $0.034 \times 0.031$  Esophagus long and 0.14 in lengths which bifurcate at the separation of 0.43 from foremost finish of body Caeca expands upto center degree of ovary and measure 0.32 long. Excretory vesicle saccular and excretory pore terminal. Genital pore subtermiral, marginally submedian, on the dorsal side of the oral sucker Testis two, oval or subspherical post-central, somewhat covering one another, submedian, post ovarian Posterior testis littler than front and measures  $0.18 \times 0.11$  and  $0.15 \times 0.09$  foremost and back separately. Cirrus sac long, rounded sickle-molded, reaching out from genital pore to pretty much front degree of hip bone socket; encasing trilobed vesicular seminalis, cylindrical standards prostatica, encompassed by prostate organ cells; restricted long,

cylindrical ejaculatory conduit and cirrus. Ovary oral or three-sided pretesticular, submedian, central, marginally right side of the body and measures  $0.06 \times 0.05$  Receptaculum seminis long, pear-formed lying among ovary and front testis and measures  $0.122 \times 0.026$  Mehli's organ cells encompass the ootype. Uterus curled broadening upto back finish of body and rising corresponding to cirrus sac open at genital pore. Eggs oval, operculated and measures  $0.023 \times 0.010 - 0.024 \times 0.012$  Vitelline follicles huge, sporadic, reaching out from front degree of hip bone socket upto back degree of foremost testicles.

Discussion & Conclusion:-The observation found like Prevalence was maximum in the month of July and minimum in the month of December. Abundance was maximum in the month of October and minimum in the month of December. Mean intensity was maximum in the month of October and minimum in the month of December. Dominance was maximum in the month of July and minimum in the month of December. Infestation index was maximum in the month of July and minimum in the month of December which is also not satisfactory with the result of Chandra et al., (2016). It was reported that fishes have greater number and variety of helminth infection during the summer month (Dogiel et al., 1958). In further investigation it was determined that the temperature is one of the important factor controlling the trematode infection (Wang et al., 2001; Poulin and Cribb, 2002). It was explained that the development of parasites need high temperature, low rainfall and sufficient moisture (Ronde, 1993; Jadhav and Bhure, 2006). Some fishes have greater incidence and intensity of the helminth infection other than summer season (Pal, 1963; Burrough, 1978). Climatic factors affect parasites directly and the temperature amongst them in the most important single extrinsic factor which influence the parasites (Noble and Noble, 1976). The seasonal cyclicality of digenetic trematode infection in fresh water fishes is temperature dependent (Pennuquick, 1971a,b; Cannon, 1972). On seasonal aspects: Prevalence was maximum in the Rainy season and minimum in the Winter season. Abundance was maximum in the Rainy season and minimum in the Winter season. Mean intensity was maximum in the Rainy season and minimum in the Winter season. Dominance was the maximum in the Rainy season and minimum in the Winter season. Infestation index was



maximum in the Rainy season and minimum in the Winter season not similar to the result of Chandra et al., (2016).

Factors influencing the seasonal variation of trematodes are feeding habits (Pennuquick, 1971a,b) and reproductive status of the host (Evans,1977a),oxygen tention ,host species, behaviour, migration ,immunity and interaction of both biotic and abiotic factors (Chubb,1977).There is meagre positive correlation between the host size and total parasitic infection and impact of diet and feeding habits on the parasitic infection in the fish hosts were carried out by (Lugue et al.,1996 and Johnson et al.,2004).The variation in the infection with age group may be as younger fish have less capacity of feeding whereas older fish may be resistant and therefore do not allow new extra parasite burdens(LO et al.,1998;Johson et al.,2004).The temperature dependent infestation of adult digenean has been reported in *Bunodera luciopercae*(Wotten,1973a and Andrews,1977).The water temperature may be the most significant factor for understanding the seasonal dynamics of metacercaria in the fishes of mid latitude climatic zone of the world (Chubb,1979).It has reported that the incidence of infection of *Sterrharius monolecithus* and *Fastula bravichrus* remain higher in winter months with maximum in the month of september in the host fish *Hilsa ilisha* and this may due to migratory habits of fishes influencing directly the parasitic fauna of the host (Srivastava,1935; Pal,1963).

On the above findings the author knew the load of digenetic trematode parasites in *Channa striatus*. Author found that different ecological aspects shows fluctuation monthly as well as seasonally. Maximum infestation index occur in the rainy season and minimum in winter season. Diseases have an effect on the food value and market value of fish.So it is important to use different preventive measures to check the infection and also aware people about the infection. In figure one the current structure varies from all the realized species in having vitellaria follicles from the locale between intestinal bifurcation and acetabulum. The proportion of oral and ventral sucker (1:1). Prepharynx enormous. Intestinal caeca broadens upto front degree of foremost testis and receptaculum seminalis pear-formed, postero-sidelong to ovary. It further contrast M. collata and M gomia in having ovary bigger than testis, caeca upto foremost testis

while it shows similarity to the M. collata and M. chauhani in having snaked cirrus sac.The name of new species, Masenia jaunpurensis is given on locale Jaunpur.In figure two the current structure has a place with Masenia dayali Gupta, 1955 from Clarias batrachus at Saharanpur and is accounted for first time from Jaunpur with another host, Channa punctatus. It contrasts from unique depiction in having bigger throat, extention of caeca upto front degree of foremost testis and vitteline follicles from foremost degree of ventral sucker upto back degree of foremost testis.

## REFERENCES

- [1] Andrews,C.R.(1977).The biology of the parasites fauna of perch (*Perca fluviatilis* L.) from Lyn. Tegid,North wales,Ph.D.Thesis University of Liverpool.
- [2] Ashade O.O.;Osineye O.M.; Kumoye E.A.(2013). Isolation,identification and prevalence of parasites on *Oreochromis niloticus* from three selected Riverr system. J. Fish Aqua. Sci. 8:115-121 .
- [3] Athokpam V.D. & Tandon V. (2015). A survey of metacercarial infections in commonly edible fish and crab hosts . J Parasitol Dis. 2015 Sep; 39(3):429-40.
- [4] Burrough, R.J.(1978). The population biology of two species of eye fluke,*Diplostomum spathaceum* and *Tylodelphys clavata* in roach and rudd.J. Fish Biol.13;19-32.
- [5] Cannon,L.R.G.(1972).Studies on ecology of Papilliose allocreadiid trematodes of the yellow perch in Algonquin Park/.Can.J.Zool;50:1231-1239.
- [6] Chandra,K.J. &Golder M.I. (1987).Effect of helminth parasites on fresh water fishes *Nandusnandus*.Environment and Ecology,5(2):333-336.
- [7] Chandra S,Yadav N. and Saxena A.M.,(2016).An ecological aspectson digenetic trematode parasites of fresh water fishes from Uttar Pradesh.International Journal of Applied and Natural Sciences(IJANS).Vol.5,Issue 4;93-102.
- [8] Chubb,J.C.(1977).Seasonal occurance of helminth parasites in fresh water fishes.Part-I.Monogenea,Advances in Parasitology,15:133-199.
- [9] De Meeus T.;Durand P.& Renaud F.(2003).Parasites within the new phylogeny of Eucaryotes.Trends in Parasitology.18.247-51.
- [10] Dogiel,V.A.(1958). Ecology of fresh water fishes,Parasitology of Fishes,(eds-V.A.,Dogiel,G.K. Petrushevski and Y.J polyanski, Lenning University press ) Transl. by Kabata.Z.(1961)Oliver and Boyd,Edinburgh,1958,pp.1-47.
- [11] D' Silva J.; Bhuyan A.I. and Bristow G.A., (2012). Distribution of helminth parasites in size group and organs of *Hilsa shad*, *Tenulosa Ilisa*. Dhaka Univ, J.Biol. Sci, 21(1), 55-65 (2012) .
- [12] Esch G.W.&Barger,M.(2001). Downstream change in the composition of the parasite community of fishes in an Appalachian stream.Journal of Parasitology 87:250-255.
- [13] Evans,N.A.(1977a).The occurance of *Sphaerostoma bramae* (Digenea:Allocreadiidae)in the roach from the Worcester Birmingham Canal.J.Helminth;51:189-196.
- [14] FAO (2006). India,National Fishery Sector Overview : India.

- [15] FAO (2008).FAOSTAT Online Statistical Service - United Nation Food and Agriculture Organisation ( FAO ), Rome ,available from : <http://faostat.fao.org>.
- [16] Gupta,S.P.(1959).Nematode parasites of vertebrates of East Pakistan III.Camallanidae from fishes,Amphibians,Reptiles.Can.J.Zool.,87:771-779.
- [17] Jadhav,B.V. and Bhure,D.B.(2006).Population dynamics of helminth parasites in fresh water fishes of Marathwada region ( M.P.) India.Flora and Fauna :An International Research Journal,Vol.12,No.2,pp.143-148
- [18] Johnson, M.W.;Nelson,P.A.; and Dick ,T.A.,(2004). Structuring mechanism of yellow perch (*Perca flavescens*) parasitic communities : host age diet and local factors. Canadian J.Zool.82:1291-1301.
- [19] Kennedy C.R. and Guegan J.F. (1996). Parasite richness /sampling efforts/host range;the fancy three pices jigsaw puzzle.Parasitol Today.12:367-369.
- [20] Khanum H.; Begum A. and Begum S. (2011). Seasonal prevalence, Intensity and Organal distribution of Helminth parasites in *Macroglystus aculeatus*, Dhaka Uni., J.Biol. Sci, 20(2), 117-112.
- [21] Kundu I.; Bandyopadhyay P.K. & Mandal D.R. (2015). Prevalence of helminth parasites infecting *Channa punctatus* Bloch, 1793 from Nadia district of W.B. Journal of Agriculture and Veterinary Science 8:41-46.
- [22] Kuris,Armand & Lafferty,Kelvin(1994).Community Structure:Larval trematodes in Snail Hosts.Annual Review of Ecology and Systematics.25.189-271.
- [23] Leong,T.S. (1986). Seasonal occurrence of metazoan parasites of *Puntius binotatus* in irrigation canal.Pvluu Pinn.Malaysia J.Fish Biol.,28(1):9-16.
- [24] Lo,C.M.;Morand S. and Galzin,R.(1998).Parasites diversity /host age and size relationship in three Coral-reef fishes from French Polynesia.International J.of Parasitology,28:1695-1708.
- [25] Margolis L.; Esch,G.W.; Holmes,J.C.;Kuris A.M.;Schad G.A. (1982). The use of ecological terms in parasitology . J.Parasitol., 68 (1)1982 ,pp. 131-133 ©American Society of Parasitologists 1982.
- [26] Markov,G.S.(1961). Physiology of fish parasites.In parasitology of fishes(V.A.Dogiel,G.K.of Bangladesh).Bangladesh.J.Zool.1973,1:63-81.
- [27] Mishra A.;Singh Prashant; Mishra D.B. And Singh H.S. (2019). Prasence of digenetic trematode parasites of cypriniformes fish *Labeo rohita* . IJSR, vol ( 8 ) I ( 2 ) ISSN: 2319 -7064 .
- [28] Mishra Anurag; Singh Prashant; Mishra D.B. and Singh H.S. (2019). Digenean trematode parasites and cyprinid fish *Catla catla* . IJSR ,vol ( 8 ) ,I ( 5 ) ,ISSN No. 2319-7064.
- [29] Noble,E.R. and Noble ,G.A.(1976).Parasitology ;The Biology of Animal Parasites ,pp.1-556.Lea and Febiger,Philadelphia.
- [30] Pal,R.N.(1963). Observation on fluctuations in parasitization of Indian Shad.,Hilsa ilisha (Hamilton) of the Hooghly estuary.Ind.J.of Helminth.,15:119-126.
- [31] Paperna I.(1996). Parasites ,infections and diseases of fishes in Africa –an update, CIFT Technical paper no. 31, Rome ,FAO,220 p. Petrushevsky and Yu,I. polyansky eds. English translation by Z. Kobata, Oliver and Boyed.
- [32] Pennuquick,I.(1971a).Quantitative effects of three species of parasites on a population of three spined stickleback *Gasterosteus aculeatus* J.Zool.65:143-162.
- [33] (1971b) Seasonal variation in the parasitic infection in a population of three spined stickleback *Gasterosteus aculeatus*.Parasitol.,63:373-388.
- [34] Poulin,R. and Cribb,T.H.(2002).Trematode life cycles:short is sweet?Trends in Parasitology 18:176-183.Cross Ref.Pub Med,CSA.
- [35] Robert Poulin (2004).Parasites and the neutral theory of biodiversity. Ecography / Volume 27, Issue1.
- [36] Ronde,K.(1993).Ecology of marine parasites :An introduction to marine parasitology , Department of Zoology ,Unit of New England Armidale NSW 2351,Australia CAB International ISBN 8519-8458.
- [37] Sarmin Akther; Emrul Hakkani; Ashadyzaman and Moni Krishna Mohanta (2018). Prevalence of parasitic infections in the fresh water fish, *Channa punctatus* (Bloch) from Rajshahi Metropolitan , Bangladesh . Int. J. Curr. Microbiol. App. Sci. (2018) 7 (4): 3431- 3441.
- [38] Shareef P.A.A.& Abidi S.M.A. (2012). Incidence and histopathology of encysted progenetic metacercaria of *Clistostomum complanatum* ( Digenea : Clinostomidae ) in *Channa punctatus* and its development in experimental host –Asian.Pac. J.Trop. Biomed. 2:421-426.
- [39] Singh Prashant, Mishra D.B. and Singh H.S. (2018). Studies on Digenetic trematode parasites in fresh water Carnivorous fishes of Jaunpur. Int. J. of Science and Research (2018)7 (10) :2319-7064.
- [40] Singh Prashant, Mishra D.B. and Singh H.S. (2019). Occurance of digenetic trematode parasites in fresh water cat fish *Wallago attu* . Shodh Sandarsh ( UGC Approved Journal No. 41329 ),ISSN No. 2319-5908, page no.447-451.
- [41] Srivastava,H.D.(1935).New hemiurid (Trematoda) from Indian fresh water fishes Part-I-New distomes of the genus *Lecithaster* luhe 1901 from *Clupea ilisha*. Proc.Acad. Sci.U.P. India, 4:381-387.
- [42] Srivastav ,H.D.(1935).New parasites of the genus *Orientophorus* n. gen. from an Indian fresh water fish ,*Clupea ilisha*.Parasitology,(27):374-382.
- [43] Wang,GT.;Yao,WJ and Nie,P.(2001). Seasonal occurrence of *Dollfusotrema vaneyi* (Digenea: bucephalidae) metacercaria in the bull cat fish *Pseudobagrus fluvidraeo* in a reservoir of China .Disease of Aquatic Organism.44(2):127-131.
- [44] Wik. –From Wikipedia .Wooten,R.(1973a). Occurance of *Bundera luciopercae* (Digenea:Allcreadiidae) in fish from Hanningfield Reservoir,Essex.J.Helminth.,47:399-408.
- [45] Yamaguti S.(1954). Systemahelminthum vol. 1 The digenetic trematodes of vertebrates.Int. J. Sci. pub.NewYark 1575.
- [46] Zrzvy J.(2001). Ecdysozoa versus Articulata ;Clades,Artifacts and Prejudices.Journal of Zoological Systematics and Evolutionary Research.39(1):159-163.



**Tables, Graph & Figure :-** Following graph& figure were found with the help of data

| MONTH | NO OF HOST EXAMINED | NO OF HOST INFECTED | NO OF PARASITES | PREVALENCE % | ABUNDANCE % | MEAN INTENSITY | DOMINANCE |
|-------|---------------------|---------------------|-----------------|--------------|-------------|----------------|-----------|
| JAN   | 05                  | 01                  | 01              | 20.00        | 20.00       | 1.00           | 0.01      |
| FEB   | 08                  | 02                  | 03              | 25.00        | 37.50       | 1.50           | 0.03      |
| MAR   | 07                  | 01                  | 02              | 14.28        | 28.57       | 2.00           | 0.02      |
| APR   | 08                  | 03                  | 04              | 37.50        | 50.00       | 1.33           | 0.05      |
| MAY   | 14                  | 06                  | 10              | 42.85        | 71.42       | 1.66           | 0.12      |
| JUN   | 12                  | 06                  | 09              | 50.00        | 75.00       | 1.50           | 0.11      |
| JUL   | 15                  | 11                  | 17              | 73.33        | 113.33      | 1.54           | 0.22      |
| AUG   | 10                  | 05                  | 12              | 50.00        | 120.0       | 2.40           | 0.15      |
| SEP   | 06                  | 02                  | 04              | 33.33        | 66.66       | 2.00           | 0.05      |
| OCT   | 11                  | 02                  | 15              | 18.18        | 136.36      | 2.50           | 0.06      |
| NOV   | 14                  | 04                  | 09              | 28.57        | 64.28       | 2.25           | 0.11      |
| DEC   | 13                  | 01                  | 01              | 07.69        | 7.690       | 1.00           | 0.01      |

Ecological terms and *Channa striatus* (Monthly)

Table no. 1

| SEASONS | NO OF HOST EXAMINED | NO OF HOST INFECTED | NO OF PARASITES | PREVALENCE % | ABUNDANCE % | MEAN INTENSITY | DOMINANCE |
|---------|---------------------|---------------------|-----------------|--------------|-------------|----------------|-----------|
| SUMMER  | 41                  | 16                  | 25              | 39.02        | 60.97       | 1.56           | 0.32      |
| RAINY   | 42                  | 20                  | 38              | 47.61        | 90.47       | 1.90           | 0.36      |
| WINTER  | 40                  | 08                  | 14              | 20.00        | 35.00       | 1.75           | 0.18      |

Ecological terms and *Channa striatus* ( Seasonal )

Table no. 2

| MONTH | JAN  | FEB  | MAR  | APR  | MAY  | JUN  | JUL   | AUG  | SEP  | OCT  | NOV  | DEC  |
|-------|------|------|------|------|------|------|-------|------|------|------|------|------|
| I.I.  | 0.20 | 0.75 | 0.28 | 1.50 | 4.28 | 4.50 | 12.46 | 6.00 | 1.33 | 0.90 | 2.57 | 0.07 |

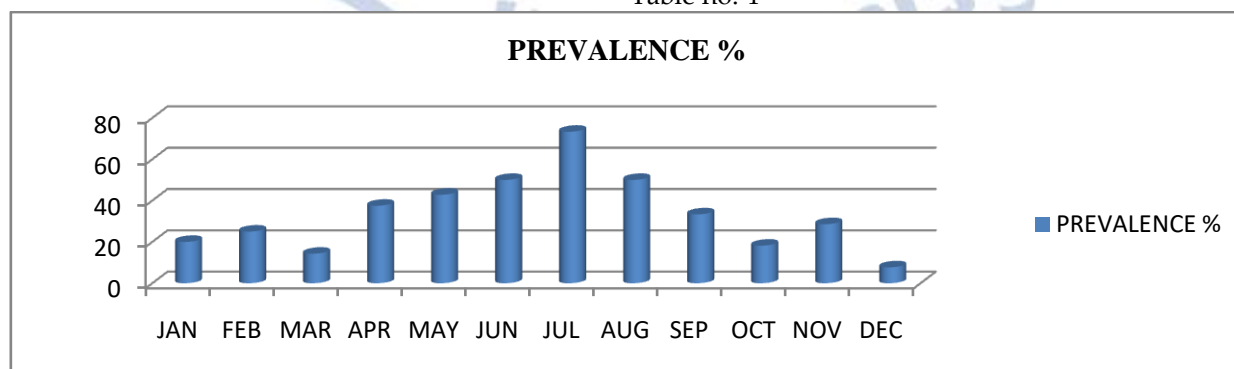
Ecological terms and *Channa striatus* ( Monthly )

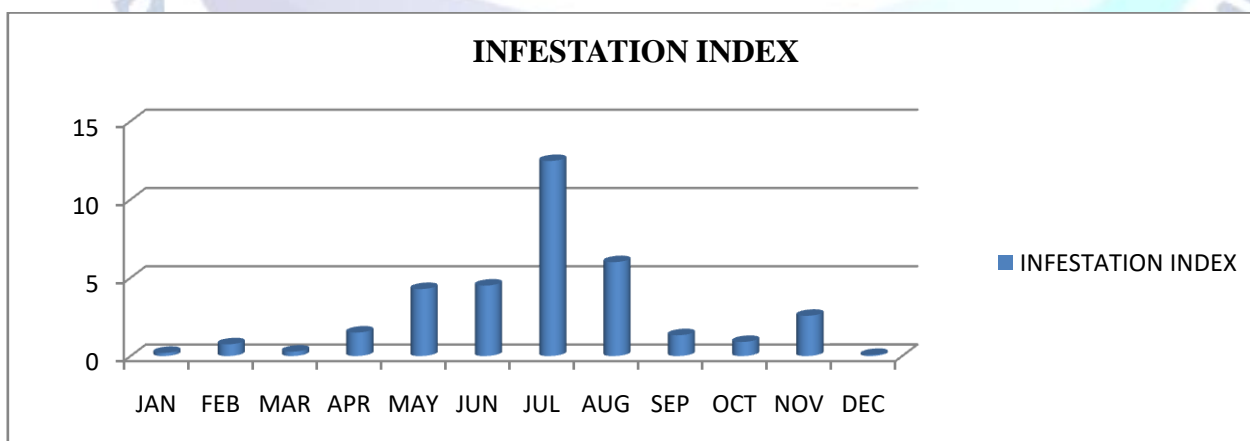
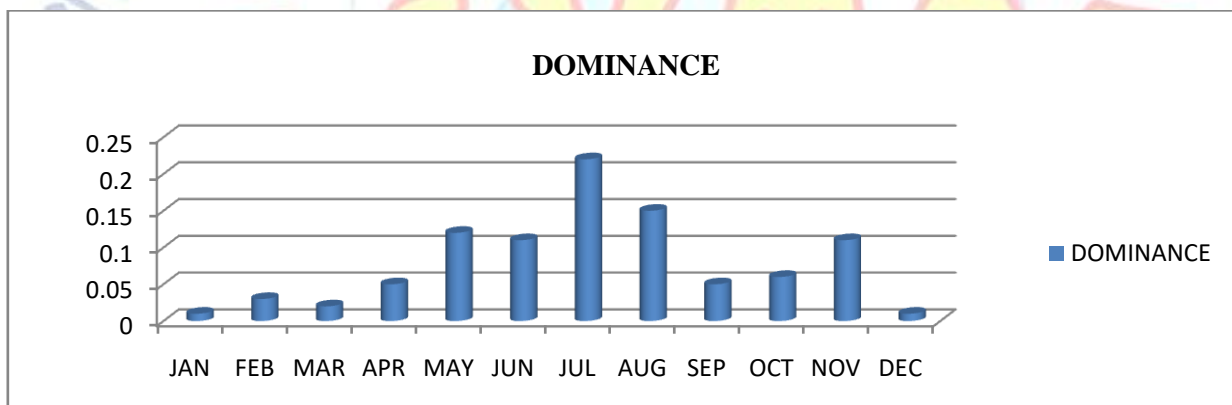
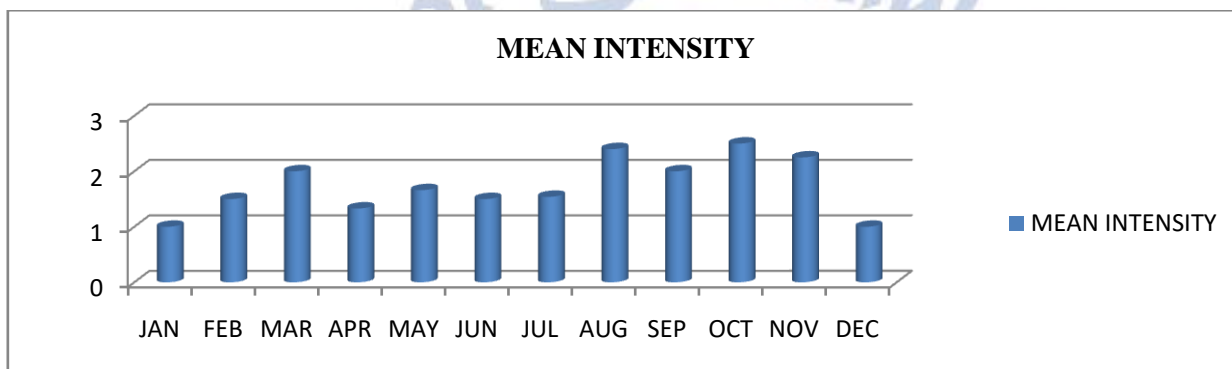
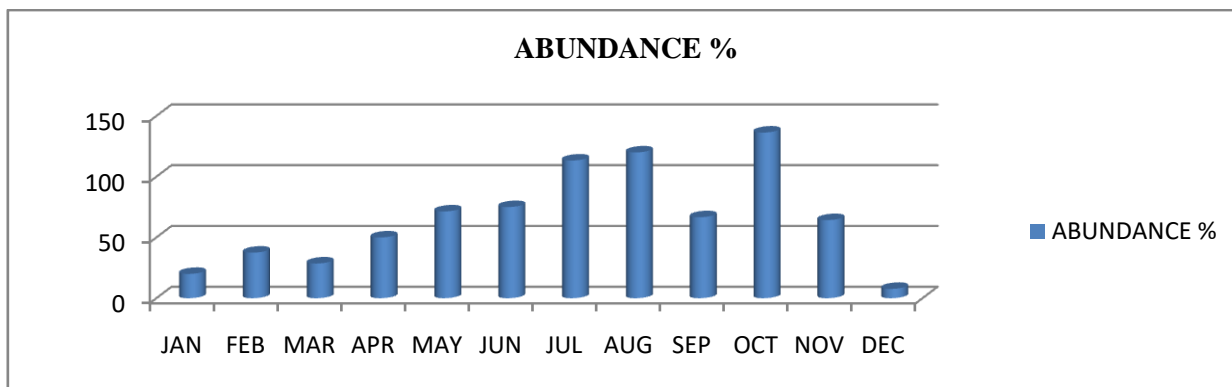
Table no. 3

| SEASONS | SUMMER | RAINY | WINTER |
|---------|--------|-------|--------|
| I.I.    | 9.75   | 18.09 | 2.80   |

Ecological terms and *Channa striata* (Seasonal)

Table no. 4

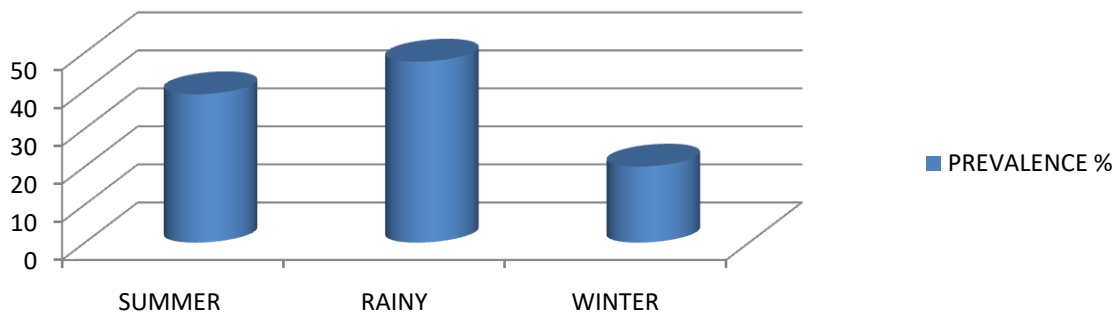




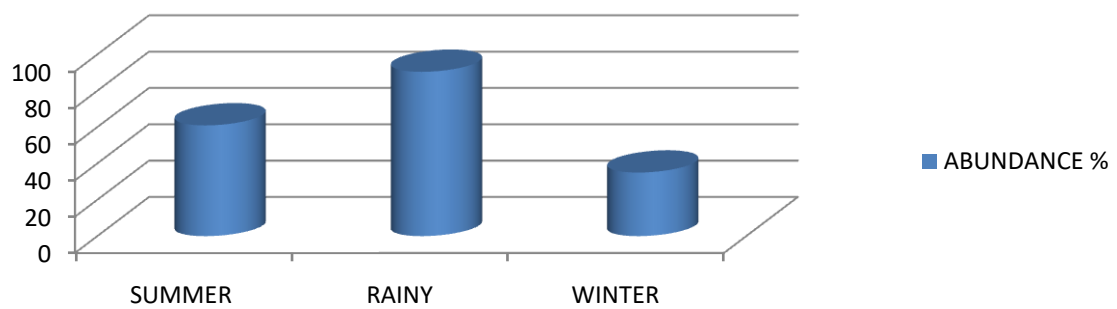
Graph, (Monthly)



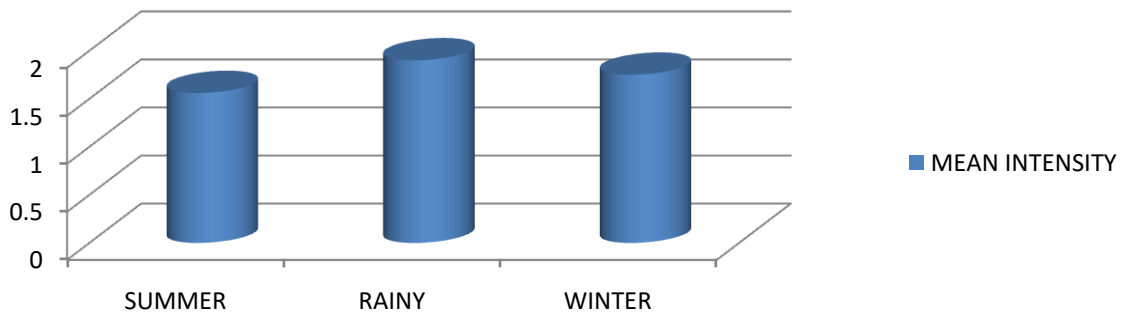
**PREVALENCE %**



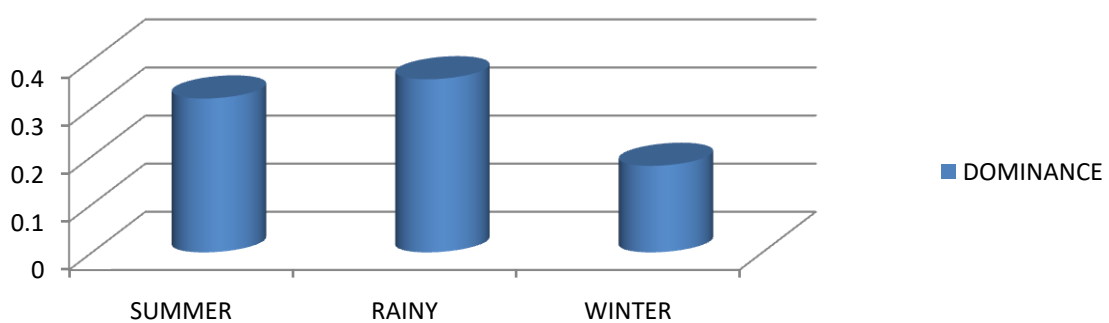
**ABUNDANCE %**

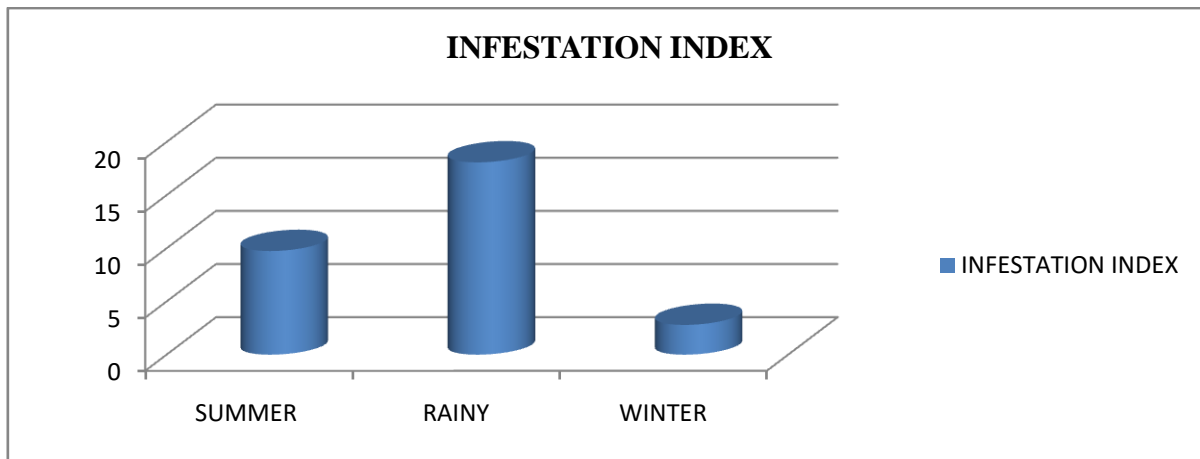


**MEAN INTENSITY**



**DOMINANCE**





Graph,(Seasonal)

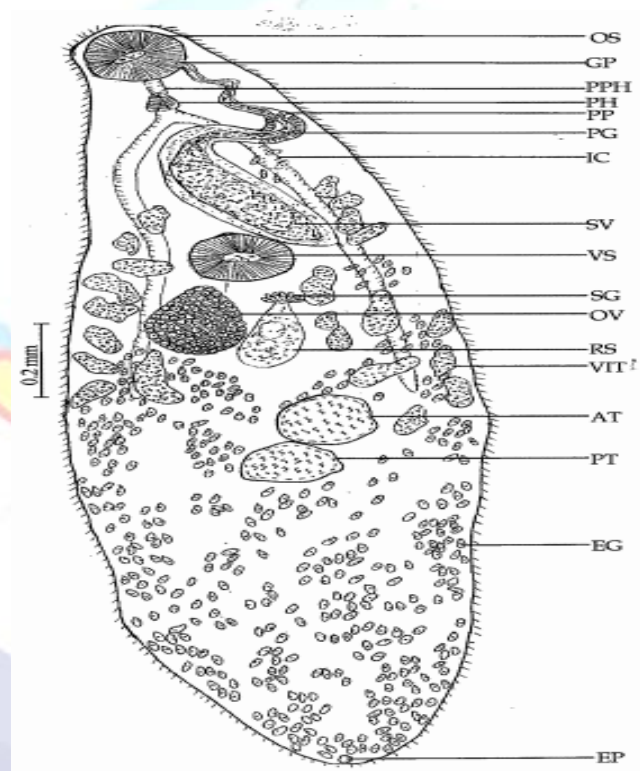


Fig 1: Maseniajaunpurensis Sp. nov.

