International Journal for Modern Trends in Science and Technology, 9(02): 237-245, 2023 Copyright © 2023 International Journal for Modern Trends in Science and Technology ISSN: 2455-3778 online DOI: https://doi.org/10.46501/IJMTST0902043

Available online at: http://www.ijmtst.com/vol9issue02.html



Green Synthesis of Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's Stabilized by a Lemon Fruit (Citrus limon) Peel Extract s. Pal For for Antioxidant Activity

Nagaraja N*, Sharanagouda, Kavya N, Deepa Pattar

Department of Physics, Sahyadri Science College, Shivamogga, Karnataka, India. *Corresponding author: E-mail: nnkadur@gmail.com

To Cite this Article

Nagaraja N, Sharanagouda, Kavya N, Deepa Pattar. Green Synthesis of Fe3O4, ZnO & Fe3O4-ZnO NP's Stabilized by a Lemon Fruit (Citrus limon) Peel Extract for Antioxidant Activity. International Journal for Modern Trends in Science and Technology 2023, 9(02), pp. 237-245. https://doi.org/10.46501/IJMTST0902043

Article Info

Received: 18 January 2022; Accepted: 20 February 2022; Published: 26 February 2023.

ABSTRACT

Now a days, nano materials have replaced all other materials in every field because of their unique physical and chemical properties, which facilitate extraordinary applications. That's why finding the most convenient way for the synthesis is very important, so in the process, researchers found the simplest way for the synthesis to be the "Green method or biological method". It is the most rapid, simple, cheap, and eco-friendly way among all the chemical and physical methods [1]. In this present work, the Fe3O4, ZnO, and Fe3O4-ZnO NPs were synthesised by the Green method using lemon fruit peel extract as a stabilizing agent [2]. The synthesised NPs were characterised by UV-VIS, FTIR, XRD, TGA, and SEM techniques. And further, the antioxidant activity of all prepared NPs [3] was studied.

Keywords: Green Synthesis: Metal Oxide NP's, Antioxidant property

1. INTRODUCTION

NPs are those whose at least one dimension should be smaller than 100nm. By virtue of their larger surface area, they have unique physical and chemical properties [4], and their nanosize facilitates cellular and molecular-level biological interactions [5]. In recent decades, nanotechnology has occupied most all fields. Especially metal oxide NPs have drawn greater interest as they have shown remarkable applications to various fields such as the medical field, pharmaceuticals, diagnosis, anticancer, antibiotics, drug delivery, cell therapy, tissue engineering, hyperthermia, etc. [6]. In agriculture fields [7], in sensing fields to sense poisonous gases [8], and as a catalyst in waste water

treatment [9]. In the research field, efforts are going on to enhance solubility, bioavailability, stability, suppress toxicity, and improve pharmacological activity [10]. Due to their wide range of applications, determining the most convenient method for the synthesis of NPs is very important. As such, we have physical, chemical, and biological methods for synthesis, but physical and chemical methods are generally expensive, require much energy, and use toxic and hazardous chemicals, the residuals of which pose a threat to the environment [11]. To overcome these limitations and disadvantages, synthesis recently introduced a biological method synthesis), which is simple, (green low-cost, eco-friendly, and requires less time and energy [12].

This method uses natural organisms (bacteria, algae, viruses, etc.) or plant extracts as they contain proteins, enzymes, amino acids, carbohydrates, alkaloids, terpenoids, tannins, saponins, phenol compounds, sugar, and vitamins. They act as stabilising, reducing, and capping agents [13] and have the capability to control the size and shape of the NPs [14].

Here, the lemon fruit peel is used for synthesis. The lemon plant can be grown in California. West Indies, Italy, Spain, Sicily, Portugal, Florida, California, Jamaica, Australia, and grown all over India, particularly in home gardens and small orchards. Lemon peel contains volatile oil (2.5%), vitamin C, hesperidin and other flavone glycosides, mucilage, pectin, and calcium oxalate [15]. The important constituents of the volatile oil are limonene (90%), citronellal, geranyl acetate, -pinene, camphene, linalool, terpineol, methyl heptenone, octyl and nonyl aldehydes, -terpinene, -pinene, neral, and geranial [16]. The peels also contain the flavonoids eriocitrin, epigenin, luteolin, chrysoeriol, quercetin, isorhamnetin, limocitrin, limocitrol, isolimocitrol, hesperidin, coumarins scopoletin and umbelliferone, sinapic acid, and -coumaric acid [17]. Lemon peel is used as a flavouring agent, perfume, stomachic, and carminative. Externally, the oil is a strong rubefacient, and if taken internally in small doses, it has stimulating and carminative properties [18].

2. Materials and Methodology :

2.1 Materials :

Lemons were collected from the local market Shivamogga, Karnataka, India. The FeCl₃.6H₂O, FeCl₂.4H₂O, $Zn(NO_3)_2$ 3H₂O and 2,2-Diphenyl-1-1-Picryllhydrazyl (DPPH) were procured from Sigma-Aldrich, India. Ethanol and acetic acid were purchased from SD-fine Chemicals, India. Double distilled water will be used throughout the experiment. Chemicals and compounds used are analytical grade and used directly as received without further purification.

2.1.1 Preparation of Lemon Fruit Peel Extract

The extract of Lemon fruit peel was obtained using a reported method. Briefly, the fruit peels were washed several times to remove dust, followed by drying at ambient temperature. A total of 10 g of the fruit peel was ground and mixed with 100 ml of double deionised water at 80°C using an oil bath under constant stirring at 250 rpm for 1 h. The crude extract solution (10 mV) was filtered with filter paper (Fioroni 601) and oven-dried at 45°C for 24 h. The dried extract powder was termed S5 and stored at 4°C for further processing.

2.1.2 Green Synthesis of Fe₃O₄ NPs by Lemon Fruit Peel Extract:

A simple co-precipitation method and the extract of Lemon fruit peel as a novel stabilising and capping agent were used to synthesise Fe₃O₄ NPs. The crude extract of Lemon fruit peel and iron salts are used as stabilisers, iron sources, and reducing agents, respectively. Four different extract solutions (wt.%) were prepared. In total, 1, 2, 5, and 10 g of the dried extract powder were added into four different beakers containing, respectively, 99, 98, 95, and 90 g of double deionised water and stirred for 15 minutes at room temperature. FeCl₃. 6H₂O, 97% (2.53 g), and FeCl₂.4H₂O, 99% (0.99 g), at a molar ratio of 2:1, respectively, were added to each solution. After that, Fruit peel extract (11.2 pH) was drop wise added into the respective solutions to adjust the pH to 11, followed by vigorous stirring for another 30 min. Finally, the samples were centrifuged three times at 14,000 rpm for 12 min, and the collected precipitates were oven-dried at 70°C. The same procedure without using the extract was carried out for the preparation of bare Fe₃O₄ NPs. The synthesised Fe₃O₄ NPs with 0, 1, 2, 5, and 10 wt.% concentrations of the peel extract were termed S0, S1, S2, S3, and S4, respectively, and the peel extract powder alone was termed S5.

2.1.3 Green Synthesis of ZnO NPs by Lemon Fruit Peel Extract:

Finely cut fruit peels were dried into a 40-gramme powder that was soaked with 100 ml of deionised water and boiled for 50 minutes until the colour of the aqueous solution changed from watery to light yellow. The mixture was then filtered to obtain aqueous leaf extract and boiled to 60–80 °C with stirring. To this was added 2 g of copper nitrate trihydrate [Zn(NO₃)₂ 3H₂O] in 100 ml of deionised water under vigorous stirring until black precipitate separated. After the formation of NPs, stop stirring and leave the precipitate to settle. Filter and repeatedly wash with deionised water and absolute ethanol for several times until pH reaches 7. Subsequently, washed material is dried at 80 °C for 16 h in an oven, followed by calcination at 500 °C for 4 h

2.1.4 Green Synthesis of ZnO NPs by Lemon Fruit Peel Extract:

Finely cut fruit peels were dried into a 40-gramme powder that was soaked with 100 ml of deionised water and boiled for 50 minutes until the colour of the aqueous solution changed from watery to light yellow. The mixture was then filtered to obtain aqueous leaf extract and boiled to 60-80 °C with stirring. To this was added 2 g of iron chloride hexahydrate [FeCl3. 6H2O] and copper nitrate trihydrate [Zn(NO₃)₂ 3H₂O] in a 1:1 molecular ratio and 100 ml of deionised water under vigorous stirring until black precipitate separated. After the formation of NPs, stop stirring and leave the precipitate to settle. Filter and repeatedly wash with deionised water and absolute ethanol for several times until pH reaches 7. Subsequently, washed material is dried at 80 °C for 16 h in an oven, followed by calcination at 500 °C for 4 h.

2.2 Ultraviolet-visible Spectroscopy (UV-Vis):

Ultraviolet-visible spectroscopy (UV-Vis) refers to absorption spectroscopy in the UV-Vis spectral region. This means it uses light in the visible and adjacent (near-UV and near-infrared) ranges. The absorption in the visible range directly affects the perceived colour of the chemicals involved. In this region of the spectrum, molecules electromagnetic undergo electronic transitions. UV-Vis spectroscopy was used to investigate the formation, stability, surface modifications, and light barrier properties (opacity, and percent transmittance) of the transparency, prepared metal oxide NPs (Shimadzu, UV-1800, USA).

2.3 Fourier Transforms in Infrared Spectroscopy:

The FT-IR spectra of the prepared NPs were acquired in the range of 4000–400 cm1 using an attenuated total refraction Fourier Transform Infrared (ATR FT-IR) spectrophotometer (Thermo Scientific, Nicolet iZ10). FT-IR functional group detection helps to analyse the structural changes that take place in the nanocomposite FLMs.

2.4 X-ray Diffraction (XRD) Analysis:

The crystalline and amorphous behaviour of the synthesised metal oxide NPs can be determined by X-ray diffraction (XRD) using a Rigaku Miniflex Diffractometer in reflection mode with Cu K radiation at an accelerating voltage of 30 kV with an operating current of 10 mA.

2.5 Thermogravimetric analysis:

The thermal stability and decomposition temperature of samples the FLM were evaluated using thermogravimetric analysis (TGA) (TA-SDT650 instruments, USA). The FLM sample (4-4.5 mg) was heated in the temperature range 25-600 °C at an incremental rate of 10 °C/min under a nitrogen atmosphere (50 ml/min). TA Instruments, made in the USA, were used to perform the differential scanning calorimetry (DSC). The 1.5-2.0 mg of the FLM sample was sealed in an aluminium crucible and heated in an inert nitrogen atmosphere (50 ml/min) from 25 to 400 °C at a heating rate of 10 °C/min.

2.6 Scanning Electron Microscopy (SEM):

TESCAN (Czech Republic) was used to examine the surface morphology of the prepared NPs. Before further treatment, the samples were covered with a thin palladium-platinum conductive coating. A sputter coating was used to produce the layer. A secondary electron detector with a 30 kV accelerating voltage was used to view the surface of the sample.

2.7 Antioxidant activity:

The antioxidant activities of the samples are measured on the basis of their free radical scavenging activity using the DPPH method. The stock solution of DPPH was prepared by dissolving 3.9432 mg of DPPH in 100 ml of methanol and storing it at 40 °C until use. 2 ml of DPPH solution was mixed with 1 ml of five different concentrations (20, 40, 60, 80, and 100 g mL-1) of the samples and standard, respectively. A mixture of 1 ml of distilled water and 2 ml of DPPH solution was used as the control. The reaction mixture was mixed, kept in the dark for 30 minutes, and incubated at room The absorbance was recorded temperature. spectrophotometrically at 517 nm. An antioxidant activity was estimated based on the percentage of DPPH radicals scavenged in the following equation:

Scavenging effect % = [control absorbance - sample absorbance] X 100

[Control absorbance]

3. Results and discussion ;

Present work demonstrated the synthesis of Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's by greener method using Lemon Fruit (Citrus limon) Peel Extract, and prepared NPs was confirmed by various techniques.

3.1.Physico-chemical characterization of Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's :

To inspect the formation of NPs and to control the reaction was collected UV-visible absorption of λ_{max} . The absorption spectra showed a distinct peak at 252nm, 254nm, 250 nm for Fe₃O₄, ZnO, Fe₃O₄-ZnO NP's [19-22]. They are also overlapped in Fig.1. The absorption spectra are distinctly separated and it's clear that, the formation of a Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's were takes place by the frut peel extraction method. It has been well established by researchers, the fruit peel contains phytoconstituents like flavonoids, polyphenols, tannin, phytosterols and many more active ingredients are responsible for the formation of NPs[23].To check the chemical composition of the prepared Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's, employed FTIR using KBr pellet method for to find stretching frequencies. The FTIR spectrum as shown in Fig.2. of Fe₃O₄, ZnO, Fe₃O₄-ZnO NP's were found characteristic peak at 3485, 3488, 3490 cm-1 are attributed to the stretching vibrations of -OH group. 2987,2990,2992cm⁻¹ are attributed to the stretching vibrations of C-H stretching and 2255,2256, 2258cm⁻¹ of -CH₂ group present in phenolic compounds of lemon peel extract [24]. The peak at 1651,1653,1656 and 1479,1483,1484 cm⁻¹ corresponds to the asymmetric and symmetric stretching of the Fe-O and Zn-O bond The band appeared at 1140.1143,1145 cm⁻¹ was assigned to the C-O stretching vibration of alcohol and ester present in the extract [25-27]. The characteristic peak at 562.565,567 cm⁻¹ was associated with the Fe-O, Zn-O stretching vibration of NPs [28]. Fig.3. represents the XRD pattern of prepared Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's, showing prominent characteristic diffraction at 24.22°,27.15°,29.34° which corresponds to the unit cell of (110). $29.4^{\circ}, 32.42^{\circ}, 32.48^{\circ}$ which corresponds to the unit cell of (111). 32.51º,32.62 º,34.45º. which corresponds to the unit cell of (111). 36.65°, 37.28°, 37.66° which corresponds to the unit cell of (202). $42.93^{\circ}, 39.52^{\circ}, 39.82^{\circ}$ which corresponds to the unit cell of (020). 48.44°,44.92°,43.55° which corresponds to the unit cell of (202). $52.08^\circ, 51.98^\circ$ 57.64° which corresponds to the unit cell of 56.07°,56.98°,61.92° which (022/31-1).corresponds to the unit cell of (113/22-0).63.38°,63.88°,65.45° which corresponds to the unit cell of (311). $71.47^{\circ}, 78.51^{\circ}, 77.13^{\circ}$ which corresponds to the unit cell of (004/22-2) crystallographic planes respectively (JCPDS: 48-15148) [29-30]. The average particle size of the Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's was found to be 17.32

nm,15.63nm and 14.61nm respectively was obtained by applying Debye-Scherer procedure [31]. The SEM image obtained for the prepared Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's was appended in Fig. 5. Fig. 6. Fig. 7. Respectively. It is observed that, the Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's have a well-defined morphology and are nearly spherical in shape. The spectrum depicted major peaks for Fe, Zn and O, which confirms the presence of Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's. This data was also in accordance with the reported Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's from Lemon fruit peel[32].The TGA analysis Fig.4.of green synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's reveals the bond breakdown and weight loss in three stages during heat operation[33]. The first phase of weigt loss occurred at 70° -120° is due to the loss of water abosorbed by NP's[33] where as the second phase at 125º-640º which could have arisen due to the breakdown of bonds and destruction of biomolecule structure[34]. And the third phase at above 700° is due to bond breakdown in NP's[35].

3.2. Antioxidant Activity study of Green synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's :

Antioxidant properties of green synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's and their activities in food and biological systems and the ability to capture free radicals were measured on the basis of free radical scavenging activity by DPPH method. To evaluate free radical inhibition (hydrogen donating) ability of synthesized NP's the DPPH radicals were used[36]. The DPPH scavenging % and half maximum inhibitory concentration (IC₅₀) of prepared NP's is as shown in Fig. 8. As the DPPH radicals ability was decreased considerably when the concentration of NP's were increased[37] (free radicals suppressing ability follws the order IC₅₀ value of NP's follows the order as ZnO> Fe₃O₄-ZnO NP's.



Fig. 1.UV-Vis Spectra of synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's.



Fig. 2. FTIR Spectra of synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's.

 $ZnO < Fe_3O_4 < Fe_3O_4$ -ZnO NP's) of the As a result the NP's have lower IC₅₀ than standard ascorbic acid[38]. The



Fig. 3. XRD Spectra of synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's.



Fig. 4. TGA Spectra of synthesized Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's .



Fig. 5. SEM Image of synthesized Fe₃O₄ NP's .



Fig. 6. SEM Image of synthesized ZnO NP's.



Fig.7. SEM Image of synthesized Fe₃O₄-ZnO NP's .

4

bonds which confirmed the alcoholic, phenolic compounds presents in lemon peel which involved in NP's formation. Thr XRD spectra confirmed the crystallographic nature of NP's. and SEM images established the well-defined morphology and nearly spherical in shape of NP's. and The average particle size of the Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's was found to be 17.32 nm,15.63nm and 14.61nm respectively which is obtained by applying Debye-Scherer procedure.TGA confirms the weight loss of NP's with various temparatures. And further the Antioxidant property

		i	onal	17.32 nm,15.63 obtained by a confirms the temparatures.	Bnm and 14.61 applying Deby weight loss And further th	Inm respectively v ve-Scherer procedu of NP's with he Antioxidant prop	which is ure.TGA various perty
$ \begin{array}{ c c c c } & (\mu gmL^1) & activity (\%) & (\mu gmL^1) & coefficient \\ \hline \\ Standard \\ Ascorbic acid & 20 & 39.281 \pm 2.120 \\ \hline \\ & 40 & 51.612 \pm 0.965 \\ \hline \\ & 60 & 70.289 \pm 3.056 \\ \hline \\ & 60 & 81.765 \pm 2.028 \\ \hline \\ & 100 & 90.878 \pm 1.909 \\ \hline \\ & 100 & 90.878 \pm 1.909 \\ \hline \\ & & & & & & & & & & & & & & & \\ \hline \\ & & & &$		Material	Concentration	Radical scavenging	IC50	Correlation	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		2	(µgmL ⁻¹)	activity (%)	(µgmL-1)	coefficient	
Ascorbic acid 40 51.612 \pm 0.965 25.74 0.989 60 70.289 \pm 3.056 80 81.765 \pm 2.028 100 90.878 \pm 1.909 20 40.856 \pm 0.970 40 52.224 \pm 1.336 60 69.888 \pm 1.270 80 72.212 \pm 1.560 100 80.145 \pm 1.418 Fe3O4 NPS 20 41.745 \pm 0.978 40 58.178 \pm 1.350 60 62.789 \pm 1.266 80 72.830 \pm 1.488 100 81.666 \pm 1.515 Fe3O4-ZnO 20 42.455 \pm 0.966 NPS 40 60.248 \pm 1.039 540 100 61.248 \pm 1.039 55.12 60 60.248 \pm 1.670 80 72.830 \pm 1.450 0.985 0.993 40 0.993 40 0.993 40 0.993 40 0.993 0.993		Standard	20	39.281 <u>+</u> 2.120	× A		
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		Ascorbic acid	40	51.612 <u>+</u> 0.965	25.7 <mark>4</mark>	0.989	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	1		60	70.289 <u>+</u> 3.056			2
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	19		80	81.765 <u>+</u> 2.028			
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			100	90.878 <u>+</u> 1.909		SN	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	4		20	<mark>40.856+</mark> 0.970			
Conclusion 60 69.888 ± 1.270 0.987 80 72.212 ± 1.560 100 80.145 ± 1.418 0.987 100 80.145 ± 1.418 100 80.145 ± 1.418 0.987 In this prese nt work, succes 40 58.178 ± 1.350 0.985 60 62.789 ± 1.266 39.25 0.985 80 72.830 ± 1.488 0.985 100 81.666 ± 1.515 0.985 80 72.830 ± 1.488 0.985 80 72.830 ± 1.488 0.985 80 72.830 ± 1.488 0.993 81.666 ± 1.515 0.993 51.2 80 60.248 ± 1.039 35.12 80 74.965 ± 1.598 0.993 80 74.965 ± 1.598 0.993	×	ZnO NPs	40	<mark>52.224</mark> + 1.336	42.85	0	
$\begin{array}{c cccc} 80 & 72.212 \pm 1.560 \\ \hline 100 & 80.145 \pm 1.418 \\ \hline 100 & 80.145 \pm 1.418 \\ \hline Fe3O4 NPs & 20 & 41.745 \pm 0.978 \\ \hline 40 & 58.178 \pm 1.350 \\ \hline 60 & 62.789 \pm 1.266 \\ \hline 80 & 72.830 \pm 1.488 \\ \hline 100 & 81.666 \pm 1.515 \\ \hline 80 & 72.830 \pm 1.488 \\ \hline 100 & 81.666 \pm 1.515 \\ \hline Fe3O4-ZnO & 20 & 42.455 \pm 0.966 \\ \hline Sfully & NPs & 40 & 60.248 \pm 1.039 \\ \hline Synth \\ esised & 80 & 74.965 \pm 1.598 \\ \hline 60 & 67.288 \pm 1.670 \\ \hline 80 & 74.965 \pm 1.598 \\ \hline \end{array} \right) 35.12 \\ 0.993 & stud den \\ \hline \end{array}$	Concl		60	69.888 <u>+</u> 1.270		0.987	
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	usion		80	72.212 <u>+</u> 1.560		•	
FesO4 NPs20 41.745 ± 0.978 978 In this prese nt work, succes 40 58.178 ± 1.350 60 62.789 ± 1.266 99.25 0.985 80 72.830 ± 1.488 100 81.666 ± 1.515 0.985 0.985 work, succesFesO4-ZnO20 42.455 ± 0.966 35.12 0.993 $studedensynthesised6067.288\pm1.67035.120.993studedenthe10083.505\pm1.4500.993studeden$			100	80.145 <u>+</u> 1.418			
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	In this	Fe ₃ O ₄ NPs	20	41.745 <u>+</u> 0.978			
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$			40	58.178 <u>+</u> 1.350			N.
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	prese	57	60	62.789 <u>+</u> 1.266	39.25	0.985	
work, succes 100 81.666 ± 1.515 Image: Constraint of the state of the	nt	0	80	72.830 <u>+</u> 1.488			
succes Fe ₃ O ₄ -ZnO 20 42.455 ± 0.966 35.12 40 60.248 ± 1.039 35.12 0.993 500 500 500 60 67.288 ± 1.670 60 67.288 ± 1.670 60.993 5100 60.993 5100 60.993 5100 600 67.288 ± 1.670 60.993 5100 600 67.288 ± 1.670 600 67.288 ± 1.670 600 67.288 ± 1.598 600 67.288 ± 1.598 600 683.505 ± 1.450 600 <	work,		100	81.666 <u>+</u> 1.515		2	
sfully NPs 40 60.248±1.039 35.12 synth 60 67.288±1.670 0.993 stude esised 80 74.965±1.598 den den	succes	Fe ₃ O ₄ -ZnO	20	42.455 <u>+</u> 0.966		0	
synth 60 67.288±1.670 0.993 stud esised 80 74.965±1.598 den the 100 83.505±1.450 not	sfully	NPs	40	60.248 <u>+</u> 1.039	35.12		
esised 80 74.965±1.598 den	synth	- C.	60	67.288 <u>+</u> 1.670		0.993	study
100 83 505+1 450	esised		80	74.965 <u>+</u> 1.598			demo
100 00.000 <u>1</u> 1.100 NStr	the		100	83.505 <u>+</u> 1.450	6		nstrat

Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's by using Lemon fruit peel extract as a stabilising agent. Thus the structural and thermal properties were examined by various advanced techniques such as UV-vis, FTIR, XRD, TGA, SEM. The UV-vissible absorption of λ_{max} peaks were showed at 252nm, 254nm, 250 nm for Fe₃O₄, ZnO & Fe₃O₄-ZnO NP's respectively. The FTIR showed the peaks for stretching vibrations of -OH, -CH, -CH2, C-O groups and also showed the asymmetric and symmetric stretching peaks for Fe-O and Zn-O

ed the more activity for Fe₃O₄-ZnO NP's than Fe₃O₄, ZnO NP's. -

Conflict of interest statement

Authors declare that they do not have any conflict of interest.

REFERENCES

[1] Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2016). A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise. Journal of advanced research, 7(1), 17-28..

- [2] Joshi, N. C., Gururani, P., & Gairola, S. P. (2022). Metal oxide nanoparticles and their nanocomposite-based materials as photocatalysts in the degradation of dyes. Biointerface Res. Appl. Chem, 12, 6557-6579.
- [3] Nadaroglu, H., Onem, H., & Alayli Gungor, A. (2017). Green synthesis of Ce2 O3 NPs and determination of its antioxidant activity. IET nanobiotechnology, 11(4), 411-419.
- [4] Yao, J., Yang, M., & Duan, Y. (2014). Chemistry, biology, and medicine of fluorescent nanomaterials and related systems: new insights into biosensing, bioimaging, genomics, diagnostics, and therapy. Chemical reviews, 114(12), 6130-6178.
- [5] Chen, V. J., & Ma, P. X. (2004). Nano-fibrous poly (L-lactic acid) scaffolds with interconnected spherical macropores. Biomaterials, 25(11), 2065-2073.
- [6] Bhatia, S., & Bhatia, S. (2016). Nanoparticles types, classification, characterization, fabrication methods and drug delivery applications. Natural Polymer Drug Delivery Systems: Nanoparticles, Plants, and Algae, 33-93.
- [7] Ali, A., Zafar, H., Zia, M., ul Haq, I., Phull, A. R., Ali, J. S., & Hussain, A. (2016). Synthesis, characterization, applications, and challenges of iron oxide nanoparticles. Nanotechnology, science and applications, 49-67.
- [8] Chatterjee, S. G., Chatterjee, S., Ray, A. K., & Chakraborty, A. K. (2015). Graphene–metal oxide nanohybrids for toxic gas sensor: A review. Sensors and Actuators B: Chemical, 221, 1170-1181.
- [9] Kumar, M., Ambika, S., Hassani, A., & Nidheesh, P. V. (2023). Waste to catalyst: role of agricultural waste in water and wastewater treatment. Science of The Total Environment, 858, 159762.
- [10] Wang, Z., Zhang, L., Zhao, J., & Xing, B. (2016). Environmental processes and toxicity of metallic nanoparticles in aquatic systems as affected by natural organic matter. Environmental Science: Nano, 3(2), 240-255.
- [11] Yuliarto, B., Septiani, N. L. W., Kaneti, Y. V., Iqbal, M., Gumilar, G., Kim, M., ... & Yamauchi, Y. (2019). Green synthesis of metal oxide nanostructures using naturally occurring compounds for energy, environmental, and bio-related applications. New Journal of Chemistry, 43(40), 15846-15856.
- [12] Devi, D., Julkapli, N. M., Sagadevan, S., & Johan, M. R. (2023). Eco-friendly green synthesis approach and evaluation of environmental and biological applications of Iron oxide nanoparticles. Inorganic Chemistry Communications, 110700.

- [13] Aslam, M., Abdullah, A. Z., & Rafatullah, M. (2021). Recent development in the green synthesis of titanium dioxide nanoparticles using plant-based biomolecules for environmental and antimicrobial applications. Journal of Industrial and Engineering Chemistry, 98, 1-16.
- [14] Hussain, I., Singh, N. B., Singh, A., Singh, H., & Singh, S.C. (2016). Green synthesis of nanoparticles and its potential application. Biotechnology letters, 38, 545-560.
- [15] Alamgir, A. N. M., & Alamgir, A. N. M. (2017). Pharmacognostical Botany: Classification of medicinal and aromatic plants (MAPs), botanical taxonomy, morphology, and anatomy of drug plants. Therapeutic Use of Medicinal Plants and Their Extracts: Volume 1: Pharmacognosy, 177-293.
- [16] Lim, T. K., & Lim, T. K. (2013). Elettaria cardamomum. Edible medicinal and non-medicinal plants: volume 5, fruits, 818-827.
- [17] Tanvashi, A., Kare, D., & Maynale, V. Use of Natural Herbs in Management of Covid19 Pandemic.
- [18] Selvam, A. (2008). Inventory of vegetable crude drug samples housed in botanical survey of India, Howrah. Pharmacognosy Reviews, 2(3), 61.
- [19] Lalthlengliani, J., Gurung, J., & Pulikkal, A. K. (2022). Solubilization of aqueous-insoluble phenothiazine drug in TX-100 micellar solution and interactions of cationic/anionic surfactants with phenothiazine–TX-100 system. Journal of Molecular Liquids, 354, 118823.
- [20] Tabibiazar, M., Mohammadifar, M. A., Roufegarinejad, L., Ghorbani, M., Hashemi, M., & Hamishehkar, H. (2019). Improvement in dispersibility, stability and antioxidant activity of resveratrol using a colloidal nanodispersion of BSA-resveratrol. Food Bioscience, 27, 46-53.
- [21] Sharma, H., Sapkota, H. P., & Dangi, N. B. (2021). A brief review of analytical methods for the estimation of allopurinol in pharmaceutical formulation and biological matrices. International Journal of Analytical Chemistry, 2021, 1-12.
- [22] Anita, M. (2015). Copper (II), Nickel (II), Cobalt (II), and Iron (II) complexes of conjugated organic ligands as potential dye-sensitised solar cell materials/Anita Marlina (Doctoral dissertation, Universiti Malaya).
- [23] Aslam, M., Abdullah, A. Z., & Rafatullah, M. (2021). Recent development in the green synthesis of titanium dioxide nanoparticles using plant-based biomolecules for environmental and antimicrobial applications. Journal of Industrial and Engineering Chemistry, 98, 1-16.
- [24] Choi, M. Y., Chai, C., Park, J. H., Lim, J., Lee, J., & Kwon, S. W. (2011). Effects of storage period and heat treatment on phenolic compound composition in dried Citrus peels (Chenpi) and discrimination of Chenpi with different storage periods through targeted metabolomic study

using HPLC-DAD analysis. Journal of Pharmaceutical and Biomedical Analysis, 54(4), 638-645.

- [25] Gök, A., İsmail Kirbaşlar, Ş., & Gülay Kirbaşlar, F. (2015). Comparison of lemon oil composition after using different extraction methods. Journal of essential oil research, 27(1), 17-22.
- [26] Kalapathy, U., & Proctor, A. (2001). Effect of acid extraction and alcohol precipitation conditions on the yield and purity of soy hull pectin. Food chemistry, 73(4), 393-396.
- [27] El-Nawawi, S. A., & Heikal, Y. A. (1995). Production of a low ester pectin by de-esterification of high ester citrus pectin. Carbohydrate Polymers, 27(3), 191-195.
- [28] Ahadpour Shal, A., & Jafari, A. (2014). Study of structural and magnetic properties of superparamagnetic Fe 3 O 4–ZnO core–shell nanoparticles. Journal of Superconductivity and Novel Magnetism, 27, 1531-1538.
- [29] Huerta-Aguilar, C. A., Ramírez-Alejandre, A. A., Thangarasu, P., Arenas-Alatorre, J. A., Reyes-Dominguez, I. A., & de la Luz Corea, M. (2019). Crystal phase induced band gap energy enhancing the photo-catalytic properties of Zn–Fe 2 O 4/Au NPs: experimental and theoretical studies. Catalysis Science & Technology, 9(12), 3066-3080.
- [30] Marand, Z. R., Farimani, M. H. R., & Shahtahmasebi, N. (2014). Study of magnetic and structural and optical properties of Zn doped Fe3O4 nanoparticles synthesized by co-precipitation method for biomedical application. Akush. Ginekol.(Sofiia), 15, 238-47.
- [31] Wang, J., Yang, J., Li, X., Wang, D., Wei, B., Song, H., ... & Fu, S. (2016). Preparation and photocatalytic properties of magnetically reusable Fe3O4@ ZnO core/shell nanoparticles. Physica E: Low-dimensional Systems and Nanostructures, 75, 66-71.
- [32] Muldarisnur, M., Zulhadjri, Z., & Arief, S. Magnetic and Optical Properties of Novel Hybrid Fe3o4/Zno-C Nanocomposites.
- [33] Fernández, L., Gamallo, M., González-Gómez, M. A., Vázquez-Vázquez, C., Rivas, J., Pintado, M., & Moreira, M. T. (2019). Insight into antibiotics removal: Exploring the photocatalytic performance of a Fe3O4/ZnO nanocomposite in a novel magnetic sequential batch reactor. Journal of environmental management, 237, 595-608.
- [34] Karuppasamy, P., Kamalesh, T., Mohankumar, V., Kalam, S. A., Pandian, M. S., Ramasamy, P., ... & Rao, S. V. (2019). Synthesis, growth, structural, optical, thermal, laser damage threshold and computational perspectives of 4-nitrophenol 4-aminobenzoic acid monohydrate (4NPABA) single crystal. Journal of Molecular Structure, 1176, 254-265.
- [35] Atacan, K., & Özacar, M. (2015). Characterization and immobilization of trypsin on tannic acid modified Fe3O4

nanoparticles. Colloids and Surfaces B: Biointerfaces, 128, 227-236.

- [36] Shanmugam, C., Sivasubramanian, G., Parthasarathi, B., Baskaran, K., Balachander, R., & Parameswaran, V. R. (2016). Antimicrobial, free radical scavenging activities and catalytic oxidation of benzyl alcohol by nano-silver synthesized from the leaf extract of Aristolochia indica L.: a promenade towards sustainability. Applied Nanoscience, 6, 711-723.
- [37] Akhavan Hezaveh, T., Pourakbar, L., Rahmani, F., & Alipour, H. (2020). Effects of ZnO NPs on phenolic compounds of rapeseed seeds under salinity stress. Journal of Plant Process and Function, 8(34), 11-18.
- [38] Argade, P. A., Bhutkar, M. A., & Magdum, C. S. (2019). Albizzia lebbeck extract mediated synthesis of Zinc Oxide Nanoparticles and study of its In-vitro Anti-diabetic and Anti-oxidant activity. Asian Journal of Pharmacy and Technology, 9(2), 93-98.
- [39] Mahboubedh Ghasemian Dazmiri, Heshmatollah Alinezhad,,Zinatossadat hossaini: Green synthesis of Fe3o4/ZnO magnetic core shell NP's for antioxidant and antimicrobial activity,Applied Organometallic chemistry.2020:e5731.
- [40] Bilal Mughal, Syed Zohaib Javaid Zaidi, Xunli Zhang, Sammer UI Hassan: Review-Applied Sciences 2021, 11, 2598
- [41] Waseem Ahmad, Krishna Kumar Jaiswal & Mohd Amjad: Inorganic & Nano-Metal Chemistry 2020,10,1080.
- [42] Amir Azizi: Jornel of Inorganic & Organometallic Polymers & materials (2020) 30:3552-3561.
- [43] G Sathishkumar, V.Logeshwaran, S.Sarathbabu, Pradeep K & sivaramkrishnan: Green synthesis of magnetic Iron oxide NP's: Artificial cells, Nanomedicine & Biotechnology 2017,141,2169.
- [44] Rouhollah Heydari, Masoumesh Foroutan & Seied Mahdi Pourmortazavi: Antobacterial Activity of Fe3O4/Cu Nanocomposite (7) Chemistry Selecct 2019,4,531-535.
- [45] Sada Venkateswarlu, B.Natesh Kumar, B.Prathima K.Anita & N.V.V. Jyothi: Elsevier Physica B(2014)10,1016.
- [46] Waseem Ahmad, R. Lohmus, I. Hussainova, A. Pokropivny,S. Vlassov. Introduction to nanomaterials and nanotechnology, Ukraine: Tartu University Press. (2007) 45-100.
- [47] Somayesh Mirsadeghi, Hamed Zandavar, Mohmmad Yousefi, & Hamid Reza Rajabi: Elsevier Journel of Environmental Management270 (2020) 110831.
- [48] Shorish M.Abdullah, Kamar Kolo, & S.Mohammad Sajadi: Green Synthesis of ZnO@TiO@SiO2 & Fe3O4@SiO2 Nanocomposite Food Science & Nutrition 2020:00:1-11.

- [49] Wei-Hong Li & Ning Yang: Green Synthesis of Ag-Fe3O4 nanocomposite for Antibacterial Activity; Material Letters2015 30571-1.
- [50] Culi Xing, Pengtao, Meng Zhao, Lin Sun & MIngxue Li: Decatungstate-based nickel(II) complex coated onto modified Iron Oxide NP's: Royal Society of Chemistry, 23, 3919-3928.
- [51] Mostafa Yusefi, Kamyar Shameli, &Kameli Kuca: Green Synthesis of Iron oxide NP's for Hyperthermia & Anti cancer Activities: International Journel of Nanomedicine:2021:16 2515-2532.
- [52] Ghodrat Mahmoudi, Ebrahim Sufimahmoudi & S.Mohammad Sajadi: Bioactive metal oxide NP's from common fruit wastes: Food Science & Nutrition: 2020:001-11.

nal For

- [53] Mahnaz Mahdavi, Farideh Namvar & Mansor Bin Ahmad: Green synthesis of Iron oxide NP's using seaweed aqueous extract: Molecules 2013,18,5954-5964.
- [54] Dalia M.S.A. Salem, Mona M.Ismail & Hermine R.Z.Tadrors: Antibiofilm activity of Gren Iron oxide NP's: Egyptian Journel of Aquatic Research:46(2020) 333-339.
- [55] D.Arockia Jency, K.sathyavathi, M.Umadevi & R.Parimaladevi: Bioactivity of Fe3O4-Au Nanocomposite: Materials Letters: 2019(19) 31426-0.

oouus puu asuaiss